Effect of different mixtures of some bioagents and Rhizobium phaseoli on bean damping-off under field condition

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ABSTRACT—Bean is considered a very common vegetable crop in many countries worldwide. Beans are used as green pods or as dry beans. Egypt cultivates beans either for local consumption or for exportation especially during winter and spring seasons. Root rot and damping-off during early stages of bean growth, cause great losses for growers. Growers usually use highly toxic chemical fungicides to protect their investment and get rid of damping-off problem. Using these toxic chemicals led to rejection of exported shipment. The present work offers biological control as a substitute for the toxic chemicals. In this work three different biocontrol mixtures were used. These mixtures consist of Trichoderma spp. isolates (T1+T2) or mixture of Bacillus subtilis isolates (B1+B2) or combination between T. harzianum isolate plus B. subtilis isolate (B2+T2) to illustrate their effect on plant protection against damping-off and also their side effect on beneficial bacterium Rhizobium phaseoli. Data obtained showed that the mixture of the different bioagents either formulated as powder or suspension were varied in their effect on controlling damping-off disease in beans under field condition. A mixture consists of B. subtilis+ T. harzianum (B2+T2) either in form of powder or suspension showed the highest effect in disease control compared with control treatment and the other two mixtures (T1+T2) and (B1+B2). Using of the mixture (B2+T2) as powder or suspension led to increase in fresh and dry weight of shoot and root system of treated plants compared with all other treatments. Using of the same mixture (B2+T2) also led to an increase in the number of pods/plant, yield and percentage of dry weight/ 100(g) pods. Regarding effect of biological treatment on chemical components of the treated plants, it was clear that using the mixture of (B2+T2) led to clear increase in free phenols, total phenols, reduced sugars, amount of chlorophyll in addition to percentage of protein compared with control treatment and the other used mixtures(T1+T2) or (B1+B2). When Rh. phaseoli was considered in plant protection process, obtained data indicated that Rh.phaseoli alone has very slight effect against incidence of damping-off disease. This slight effect increased in presence of any used biological mixtures. The highest synergistic effect was recorded when Rh. phaseoli was in combination with the mixture (B2+T2). Obtained results also revealed that the presence of Rh. phaseoli with biocontrol mixtures led to changes in some chemical components in the treated plants compared with the control and the other mixtures and Rh.phaseoli alone. The highest increase in these components (free phenol- total phenol- reduced sugars- chlorophyll- protein) was noticed when Rh.phaseoli was used in presence of (B2+T2) mixture. Effects of adding Rh.phaseoli with biocontrol mixtures on yield were also studied under field conditions. Data obtained indicated that all mixtures of biocontrol agents in combination with Rh.phaseoli led to increase number of rhizobium nodules, fresh and dry weight of roots and shoots, in addition to increase in number of pods/plant, yield/plant and weight of 100 pods.

Key words- Bacillus subtilis, bean, Biocontrol agents, damping-off, Rhizobium phaseoli, Trichoderma harzianum.

1 INTRODUCTION

Bean is considered as a very important vegetable

crop worldwide. Bean is consumed as green pods or dry bean seeds. In year 2012 about 115,750 Feddan were cultivated with different bean varieties. This area produced about 251,279 ton of green beans and 69,486 ton of dry seeds (Agriculture Statistics, 2012). Exported amount of green bean during 2013 reached about 35,881 ton with cash value about 392,881 L.E. (Anonymous, 2013). This shows the economic importance of bean as vegetable crop either for local consumption or exportation. Beans are attacked by numerous diseases. These diseases can attack all plant parts i.e. roots, leaves, stems and pods. (Graham et al., 1997 and Mukankusi *et al.*, 2010). Soil borne pathogens play a very important role in quantity and quality of bean production process. Rhizoctonia solani, Fusarium solani, Sclerotium rolfsii are considered the most aggressive and destructive pathogens causing high losses in bean under field conditions (Abeysinghe, 2007; Abd-El-Khair et al., 2011 and Pena et al., 2013). In general using chemical fugicides in vegetable production lead to toxic residues in the produced vegetable product. These residues have negative effects on

consumer health and also our environment and biological balance in the soil. Countries import Egyptian green bean ask for nearly free of chemicals in the imported beans. Presence of these chemicals in bean may be lead to rejection of all shipment. Biological control of soil borne pathogens attack bean were successfully tested and used before (Elad *et al.*,1980; Estevez de Jensen *et al.*, 2002 and Ahmadzadeh *et al.*,2009).

This work illustrates the efficacy of some bioagents, *i.e.* (*Bacillus subtilis* (Schleifer, 2001) and *Trichoderma* spp. (Rifai, 1969) in presence or absence of *Rh. phaseoli* on controlling these pathogens using different formula to figure out correlation between formula and efficacy of the treatment. Also, studying the effect of combination between the bioagents and *Rh. phaseoli*, as beneficial microorganism in the soil. The work was expanded to evaluate the effect of *Rh. phaseoli*, on protein content and fresh and dry weight of treated plants.

2 Materials and Methods

In all field experiments, unless otherwise indicated, experiments were carried out during season 2011/2012 at farm located at Badr district Behera governorate. Natural infested soil in this farm belongs to light sandy soil with alkaline reaction pH 7.5. Paulista bean cultivar was used. All experiments were designed in complete randomize plots. Five plots were used for each treatment. Each plot (4×0.6 m) was used as replicate and 30 Paulista seeds (25 cm apart) were sown in each replicate. This means 150 seeds were used for each treatment.

Preparing and use of different bioagents

Different bioagents T.viridi (T1) and T.harzianum(T2) were grown separately on Gliotoxin Fermentation medium (GFM) (Brian and Hemming, 1945) for 15 days under complete darkness condition to stimulate toxin production (Abd El-Moity, 1985). B. subtilis isolate(B1) and B. subtilis isolate(B2) were grown separately on Nutrient glucose (NG) medium (Dowson, 1957) for 2 days. Different bioagents were blended in electric blender for 2 minutes. Suspension of different biocontrol agents was prepared as powder using Talc powder as a method developed by Abd El Moity (1985). The same antagonists were also prepared as suspension. Numbers of colony forming units (cfu) per each one gram of prepared powder or per each ml of prepared suspension of different antagonists were adjusted to be contain 30 x106 cfu/ 1ml or 1 g of formula. Rhizobium inoculum was prepared by inoculating pure identified Rhizobium phaseoli isolate on yeast mannitol(YM) medium (Allen, 1959) for 3 days at 25°C. Obtained culture was adjusted, to be contain 30 x 10⁶ cfu/ml. Adjusted Rh. phaseoli suspension was used, as suspension at the rate of 1L/ 100 L water as soil drench after one week from sowing time.

Antagonistic mixtures were prepared as powder or liquid suspension. Powder and suspension for were used as seed coating at the rate of 10 g or 10 ml/1 kg of seeds. Plots with only bean seeds and pathogenic inocula, without antagonists, were act as control treatment. All treatments received the same amount of fertilizers and irrigation water.

Collecting data and statistical analysis

Pre- emergence damping-off was recorded 15 days after sowing. Whereas post-emergence was recorded 30 days after sowing. Percentage of damping-off was calculated according to next form:

Percentage of damping-off = $\frac{N1-N2}{N1} \times 100$

N1 = Number of sown seeds

N2 = Number of seedlings

All data were subjected to statistical analysis using the standard procedures including general linear model (GLM) available in SAS (1996). Duncan's multiple- range test was also used to illustrate significances among different means.

Effect of using different bioagents on controlling damping-off disease in beans under field conditions

The efficacy of the mixture of different biocontrol agents in form of powder or suspension, this experiment was conducted.

Paulista bean seeds were sown in natural infested soil, as described before, different antagonistic mixtures were used

in two forms as powder at the rate of 10 g/ 1kg or as suspension at the rate of 10 ml/ 1kg of bean seeds as seed coating treatment. Plots only received pathogens without biocontrol treatments were act as control treatment. Data were collected 15 and 30 days after sowing for preemergence and post-emergence damping-off. Survived plants were recorded 45 days after sowing. Data were subjected to statistical analysis as mentioned before.

Effect of using different bioagents on some plant characteristics of bean

To confirm that there are no negative effect due to using any of the used antagonists either in powder or suspension forms, yield component, fresh and dry weight (g)/plant, number of pods, yield/plant and dry matter/100g pods were determined in mature plants. Fresh weight was, also, determined using calibrated digital balance. Dry weight was determined by drying plant parts on 70° C for 5 days.

Effect of using different bioagents on some chemical components of bean plants

The effect of different bioagents on the chemical composition of the treated plants was carried out. Paulista mature plants were collected randomly from each treatment. Collected plants were cleaned up and extracted to determine free, conjugated and total phenols according to the method developed by Simons and Ross (1971). Amount of reduced and non reduced sugars were also determined using the method mentioned by Thomas and Dutcher (1924). Protein content in addition to amount of chlorophyll were also assessed using the methods described by Bradford (1976) and Nornai (1982), respectively.

Reaction between *Rhizobium phaseoli* and mixture of the bioagents

Effect of combination between biocontrol mixture as suspension and endo-nitrogen fixer bacterium *Rh. phaseoli* on damping-off disease incidence and on capacity of *Rh. phaseoli* in nitrogen fixation, expressed as percentage of protein content, in the treated plants were studied. The following two experiments were designed and conducted to illustrate the effect of the presence of *Rh. phaseoli* on capacity of different bioagents on controlling damping-off. On the other hand, the effect of different bioagents on capacity and activity of *Rh. phaseoli* expressed as protein content in different plants received different biocontrol treatment. *Rh. phaseoli* was grown on yeast mannitol (YM) medium for 3 days at 25°c. *Rh. phaseoli* was added to the soil at the rate of 1L/ 100 L water as soil drench after one week from sowing time. Bioagent mixtures were added as described before.

Effect of using different bioagents in presence or absence of *Rh. phaseoli* on incidence of damping-off diseases

The different mixtures of the bioagents were used on bean seeds alone or in combination with *Rh. phaseoli* inoculum to test their effect on the incidence of damping-off. Data of preand post-emergence damping-off in addition to percentage of the survived plants were determined as described before to figure out if any negative or positive effect due to the presence of *Rh.phaseoli* on the efficacy of the bioagents.

Effect of using the mixture of biofertilizer bacterium *Rh.Phaseoli* with the different bioagents on some chemical components of bean plants

To illustrate and compare effect of *Rh.phaseoli* on protein content of the treated plants compared with the effect of the same nitrogen fixer bacterium in combination with different bioagent mixtures, this experiment was carried out. Bean seeds Paulista variety, were treated separately using different biocontrol mixtures (B1+B2), (T1+T2) or (B2+T2) at the rate of 10 ml/1kg of bean seeds, at sowing time. *Rh.phaseoli* was added as soil drench at the rate 1:100 ml water after one week from sowing time. Plots only received *Rh. phaseoli*, without biocontrol agents, in addition another group of plots without any treatment, neither biocontrol agents nor *Rh. phaseoli* were used as control treatments. All plots received the same amount of fertilizers and irrigation water.

Effect of using the mixture of biofertilizer bacterium *Rh. phaseoli* with the different bioagents on bean plant characteristics

To compare the effect of *Rh. phaseoli* alone with the effect of *Rh. phaseoli* in combination with different bioagent mixtures on characteristics of the treated plants, this experiment was carried out. Another aim of this experiment was to be sure that using different bioagent mixtures have no negative effect against the beneficial bacterium *Rh. phaseoli*.

Bean seeds were treated with different bioagent mixtures, separately, and then *Rh. phaseoli* was added to soil at the rate of 1L / 100 L water as soil drench after one week from sowing time. Seeds were sown in plots at the rate of 30 seeds per plot. Five plots were used for each treatment. At ripening stages, randomize plants were collected from each treatment to determine fresh weight of root and shoot system. Roots and shoots were then dried using electric oven at 70°C for 5 days and dry weight was determined. Yield and number of pods/plant were also determined in addition to dry matter of 100 g of pods. Data were statistically analyzed and means were compared using Duncan's multiple-range test.

early stages of growth causing damping-off. At the same time to be sure about if there is any side effect of these bioagent treatments on *Rhizobium* as beneficial soil borne biofertilizer.

Data presented in Table (1) indicate that using the mixture of *T.viride+T.harzianum* (T1+T2) or mixture of both *B. subtilis* isolates (B1+B2) or the combination between B2+T2, either formulated as powder or suspension led to clear significant reduction in damping-off disease in beans compare with control treatment. In all cases, when bioagents were used, no diseased plants were recorded during post emergence stage compare with 20% post- emergence damping-off in control treatment. Percentage of survived plants was increased in all biocontrol treatments compare with control. Table (1) also shows that the mixture that contained *B. subtilis* isolate2 + *T. harzianum* isolate2 (B2+T2) resulted in the best effect on reducing damping-off , which only 16.7% damping-off was recorded either when this mixture was used as powder or suspension.

When the mixture of both *B. subtilis* isolates was used (B1+B2), significant different was noticed between using the same mixture as powder or suspension, where 20% damping-off was recorded in case of using the suspension, meanwhile, 30% was recorded when the same mixture was used as powder. (Abo-Shosha, 2006).

These data could be explained in the light of facts that T. harzianum can grow very fast and produces Gliotoxin and Trichodermin (Abd-El-Moity, 1981, Abd El-Moity, 1985, Balode, 2010, Abdollahi et al., 2012), whereas B. subtilis produces 66 different antibiotics (Ferreira et al.,1991). The combination between the two different bioagents increase spectrum and duration of the establishment of the antagonists in the rhizosphere of the treated plants. This reflected directly as an increase in the percentages of survived plants ,where 83.3% survived plants was recorded when B2+T2 was used compare with 80 and 76.7% when (B1+B2) or (T1+T2) were used as suspension, respectively. The same bioagent mixtures (B1+B2) or (T1+T2) were of less efficacy, being 70.0 and 73.4% survived plants when these bioagent mixtures were used as powder compared with 80.0 and 76.7% in case of using the same mixtures as suspension. This could be justified according to facts that the suspension can spread very quickly in the court of infection and occupied the area around root system preventing the pathogen to come close and negatively affected on the root. Antagonists as powder need some time to elaborate and spread out surround the root. As a result, suspension showed better results.

3 Results and Discussion

The aim of this work is to find out a biological method can replace using toxic chemical fungicides to protect bean plants against some soil borne pathogens attack bean at

Table 1: Effect of using different bioagents, formulated as powder or suspension, on controlling damping-off disease in beans under field conditions.

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		% Dam	ping-off					
	Pre-en	nergence	Post-er	nergence	Survived plants (%)			
Bioagents*	Suspension 10ml/kg seeds	Powder 10 g/kg seeds	Suspension 10ml/kg seeds	Powder 10 g/kg seeds	Suspension 10ml/kg seeds	Powder 10 g/kg seeds		
T_1+T_2	23.3 ^d	26.6 ^c	0.0 ^b	0.0 ^b	76.7 ^c	73.4 ^d		
B ₁ +B ₂	20.0 ^e	30.0 ^b	0.0 ^b	0.0 ^b	80.0 ⁰	70.0 ^e		
B_2+T_2	16.7 ^f	16.7 ^f	0.0 ^b	0.0 ^b	83.3ª	83.3ª		
Control	5	6.7 ^a	2	0.0 ^a	23.3 ^f			

Means with the same letter are not significantly different.

* T₁+T₂= the mixture of *T.viride*+*T.harzianum* (T1+T2), $B_1+B_2 =$ the two *B. subtilis* isolates and $B_2+T_2=B$. subtilis isolate2 + T. harzianum isolate2.

target for most of research works. To figure out effect of second rank after (B2+T2), with no significant differences, different bioagents on yield components (shoots- roots and regarding number of pods / plant, fresh and dry weight of pods) either on fresh or dry weight, this work was carried out. yield. Regarding other parameters, root, shoot weight either Data shown in Table (2) indicate that using any of the fresh or dry indicated that no or slight difference were antagonist mixture led to significant increase in all the recorded when these parameters were determined in case of determined parameters included fresh or dry weight of root, using T1+T2 or B1+B2 either as suspension or powder. shoot and pods, in addition to number of pods / plant These results can be explained in light of facts, that T. compared with control treatment. Data also revealed that the *harzianum+ B.subtilis* provide good protection for root system mixture of (B2+T2) showed the highest effect in all measured (Yobo et al., 2011). T. harzianum produces group of plant parameters compared with all the other treatments or control. growth promoting (PGP) compounds, which stimulate and No or slight significant differences were noticed in percentage improve root and shoot system and of course increase yield of dry matter when effect of different biocontrol mixtures as (Harman et al., 2004 and Elsayed, 2005). suspension were compared with effect of the same mixture in the form of powder.

High quality and quantities of the yield always the main The other two mixtures (T1+T2) and (B1+B2) came in the

Table 2: Effect of using different bioagents formulated as powder or suspension on some plant characteristics of bean
plants.

Bioagents*	Formulas	Fresh Weight (g)/plant		Dry weight(g)/plant		No. of Pods/	Yield(g)/	%Dry matter/100g
		shoots	roots	shoots	roots	- plant	plant	pods
T1+T2	suspension	85.5 [°]	4.4 ^c	17.9 ^{cd}	1.2 ^b	40 ^d	200 ^d	13.9 ^b
	powder	80.3 ^d	4.3 ^c	16.8 ^d	0.9 ^c	39 ^d	195 ^d	13.5 ^b
	suspension	85.1 ^c	4.5 ^c	18.7 ^c	1.5ª	43 ^c	215 ^c	14.0 ^b
B1+B2	powder	80.1 ^d	4.3 ^c	17.3 ^{cd}	1.0 ^{bc}	40 ^d	200 ^d	13.4 ^b
D2.T2	suspension	95.9ª	5.6ª	24.6 ^ª	1.6ª	53 ^ª	265 ^ª	16.7 ^ª
B2+T2	powder	92.3 ^b	5.1 ^b	21.1 ^b	1.2 ^b	50 ^b	250 ^b	16.0 ^a
Control		35.8 ^e	2.1 ^d	6.8 ^e	0.2 ^d	18 ^e	81 ^e	8.3 ^c

Means with the same letters are not significantly different.

* T_1+T_2 = the mixture of *T.viride+T.harzianum* (T1+T2), B_1+B_2 = the two *B. subtilis* isolates and B_2+T_2 = *B. subtilis* isolate2 + *T.* harzianum isolate2.

To figure out any physiological change due to using of the Results presented in Table (3) show that using any of tested bioagent, phenols, sugars, chlorophyll and protein bioagents led to high increase in the amount of free phenols content in treated plants were measured compare with control compared with control treatment. This could be explained by treatment.

the work of Mahmoud et al.(1995), Attia et al.(2011) and

fragments to plant led to stimulate free phenol production in are confirmed by Shoresh and Harman (2008); Akladious and treated plant as a self mechanism against pathogens. Increase Abbas (2012) and Karlidag et al.(2012). percentage of healthy plant appears as a result to increase free Results presented in Table (3) also revealed that the highest phenol in treated plants. Data also show that conjugated values of free phenols, reduced sugars, chlorophyll and phenols were reduced in all treated plants compare with protein were recorded when (B2+T2) was used in form of control treatment. Reduced sugar also was increased compare suspension. This confirm previous results were obtained in with control. This is due to increase biological activity in the same work regarding high capacity of this mixture treated plants. This increase in activity needs reduced sugar to (B2+T2) as suspension in control root rot diseases and protect be used in energy production. These data also in harmony bean roots against pathogens under test. Data in the same with (Abd-El-Khair et al., 2011).

adequate amount of nutrients and availability of needed differences in the measured parameters. energy, amount of protein and chlorophyll were increased in

Elsaved (2013). They stated that adding spores or mycelia treated plants compare with control treatment. These results

Table also show that the effects of other two mixtures (T1+T2) As a direct result for healthy root system and absorption of or (B1+B2) were very close to each others with slight

Table 3: Effect of using different bioagents formulated as powder or suspension on some chemical components in treated plants.

Treatments*		Phenols (Mg/g)			Sugars	(Mg/g)	Chlorophyll	Protein
		Free Conj Total Reducing Non-red.		(Mg/g)	(Mg/g)			
S** T1+T2		6.5	1.9	8.4	4.1	0.9	16.1	7.0
11712	P***	6.2	1.6	7.8	3.8	0.9	16.0	6.9
B1+B2 P	S	6.5	2.1	8.6	4.02	0.4	15.9	7.1
	Р	6.1	1.4	7.5	3.5	1.1	15.3	6.8
B2+T2	S	9.9	5.0	14.9	4.7	1.4	17.0	7.9
DZ+12	Р	8.2	3.4	11.6	4.5	1.6	17.0	7.3
Control		4.8	7.5	12.3	0.9	1.3	9.0	2.1

* T₁+T₂= the mixture of *T.viride*+*T.harzianum* (T1+T2), $B_1+B_2 =$ the two *B. subtilis* isolates and $B_2+T_2=B$. subtilis isolate2 + *T. harzianum* isolate2 , **suspension and *** powder.

Effect of combination between the tested bioagents and Rh.phaseoli on incidence of damping-off. Rh.phaseoli is a beneficial bacterium for bean plants. The present experiment was conducted to study effect of the interaction between the tested bioagents and Rh.phaseoli on the incidence of damping-off.

Data in Table (4) indicate that all the tested bioagent mixtures either at the presence or absence of Rh.phaseoli led to significant reduction in incidence of damping-off. Rh.phaseoli alone shows very slight effect in disease control and 43.3% pre-emergence damping-off was recorded compare with 56.7 % in control treatment. Using different bioagents in presence of *Rh.phaseoli* led to increase efficacy of the biocontrol treatment. When these bioagents were used in combination with Rh.phaseoli, the percentages of damping-off were reduced to be 13.3, 16.7 for Rh+B1+B2 and Rh+T1+T2, respectively, compared with 43.3% in case of using Rh.phaseoli alone. The highest reduction in damping-off was recorded to be 6.7% when mixture of Rh+B2+T2 was used. This may be due to the presence of Rh.phaseoli increased nitrogen uptake by the treated plants and in the rhizosphere. This increase act as stimulator to other antagonistic microorganisms to grow and spread out within court of infection preventing from any other pathogens to invade plant tissue.

Table 4: Effect of using different bioagents as suspension in presence or absence of Rh.phaseoli on incidence of bean damping-off diseases.

	% Dampin						
Treatments*	Pre-emergence	Post-emergence	Survived plants (%)				
Rh+T1+T2	16.7 ^d	0.0 ^b	83.3 ^c				
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Rh+B1+B2		13.3 ^d		0.0 ^b		86.7 ^b		
Rh+B2+T2		6.7 ^e		0.0 ^b		93.3 ^ª		
Rh.phaseoli		43.3 ^b		0.0 ^b		56.7 ^d		
Control		56.7 ^ª		20.0 ^a		23.3 ^e		
 Means	with	the	same	letters	are	not	significantly	different.

* Rh+T1+T2= the mixture of Rh.phaseoli +T.viride+T.harzianum, Rh+B1+B2 = Rh.phaseoli + the two B. subtilis isolates and Rh+ B₂+T₂= Rh.phaseoli+B. subtilis isolate2 + T. harzianum isolate2.

used biocontrol agents against Rh.phaseoli as beneficial correlated with protein and chlorophyll contents. This means bacterium, this experiment was carried out. Data obtained that plants with high amount of protein and chlorophyll show were tabulated in table (5). Data in table (5) show that high amount in phenols and reduced sugars and vise versa. capacity of Rh.phaseoli on protein production (nitrogen These data can be explained in the light of facts that fixation) was increased at presence of any used antagonistic Rh.phaseoli penetrates root system and works inside plant, far mixtures. Plants received *Rh.phaseoli*+B2+T2 showed the away from any outside root effect where different antagonists highest amount of protein (8.51 mg/g) and chlorophyll (18.3 are working (Elkoca et al., 2010). Also good healthy root mg/g) compare with plants received Rh.phaseoli alone (3.4, system means increase root exudates which stimulate and 12.1 mg/g) or the other two antagonistic mixtures. The other increase number of Rh. phaseoli consequently increase number two mixtures showed very close results to each others in their of nodules and percentage of protein. effect regarding all measured parameters. As noticed before

To be sure that there is no antagonistic effect from different amount of free phenols, reduced sugars were also positively

Table 5: Effect of using the mixture of biofertilizer, bacterium <i>Rh. phaseoli</i> , and different bioagents on some chemical
components of bean plants.

Bioagents*	Phenols (Mg/g)			Sugars	(Mg/g)	Chlorophyll	Protein	
Bioagenito	Free	Conj	Total	Reducing	Non-red.	(Mg/g)	(Mg/g)	
Rh+T1+T2	11.0	2.3	13.3	5.3	0.8	17.0	7.6	
Rh+B1+B2	11.7	2.0	13.7	5.4	0.7	17.4	7.8	
Rh+B2+T2	13.9	1.8	15.7	6.1	0.7	18.3	8.5	
Rh.phaseoli	9.5	2.5	12.0	2.8	0.5	12.1	3.4	
Control	4.8	7.5	12.3	0.9	1.3	9.0	2.1	

* Rh+T1+T2= the mixture of Rh.phaseoli +T.viride+T.harzianum, Rh+B1+B2 = Rh.phaseoli + the two B. subtilis isolates and Rh+ B₂+T₂= Rh.phaseoli+B. subtilis isolate2 + T. harzianum isolate2.

Different bioagents were used in presence of *Rh.phaseoli* to be sure that no negative effect can be occurred due to biological treatment. To confirm this effect, plant weights either on fresh or dry basis were determined in addition to yield component (No. of Pods/plant, yield(g)/plant, and % dry matter/100 g pods).

Data in Table (6) show that use any of biocontrol preparation led to improve in capacity of Rh. phaseoli. This improvement was reflected on plant parameters either fresh or dry weights of treated plants in addition to yield component. Rh.phaseoli+(B2+T2) was the best combination. The highest fresh weight of shoot and root were reached 119.5 g and 7.7 g /plant compare with only 35.8g and 3.5 g/plant for shoots and roots in control treatment. When Rh. phaseoli was used alone without any biocontrol treatment only 78.5 and 4.2 g/plant were recorded for fresh shoots and roots, respectively. The same results were obtained when shoots and roots were determined on dry basis and compare with control or only Rh. phaseoli treatments. As logic, healthy good shoot and root systems reflects in increase the number of pods/plant, yield (g)/plant and % dry matter/100 g pods were increased.

In Rh.phaseoli+B2+T2 mean numbers of pods/plant reach 78 pods/plant compare with only 18 in control and 40 in only Rh.phaseoli treatment. Yield and percentage dry matter also increased compare with any other treatment. This work confirms results obtained by Estevez de Jensen et al.(2002); Abd-El-Moneim et al.(2006) and Parikka et al.(2009).

Results can be explained as follow; using effective bioagents protect root system and allow these roots to grow and establish and absorb high amount of NPK converted in plants to green leaves and increase number of pods and yield. Other two mixtures show different effects.

Rh.phaseoli+B1+B2 show slight better effect compare with Rh.phaseoli+T1+T2 and number of pods reach 64/plant compare with 60/plant in Rh.phaseoli+T1+T2 where yield in Rh.phaseoli+B1+B2 reach 320.3 g/plant compare with 300 g/plant in *Rh.phaseoli*+T1+T2. Also percentage of dry matter was increased from 16.9 in case *Rh.phaseoli*+T1+T2 to be 17.6 in *Rh.phaseoli*+B1+B2.

Table 6: Effect of the mixture of the biofertilizer bacterium	Rh. phaseoli and different bio	oagents on bean plant characteri	stics.

Bioagents*	No. of nodules/	Fresh weight(g)/plant		Dry weight(g)/plant		No.of pods/plant	Yield(g)/ plant	%Dry matter/100g
	plant	nt shoots r		oot shoots root		pouo, pian	plant	pods
Rh+T1+T2	25 [°]	91.6 ^c	6.5 ^b	19.9 ^c	2.5 ^b	60 ^b	300.0 ^c	16.9 ^b
Rh+B1+B2	30 ^b	97.1 ^b	5.8 ^c	23.5 ^b	2.9 ^ª	64 ^b	320.3 ^b	17.6 ^b
Rh+B2+T2	38 ^ª	119.5 ^ª	7.7 ^a	26.6ª	3.0 ^a	78 ^ª	395.0ª	19.3 ^ª
Rh.phaseoli	18 ^d	78.5 ^d	4.2 ^d	15.0 ^d	1.3 ^c	40 ^c	195.7 ^d	11.4 ^c
Control	0 ^e	35.8 ^e	3.5 ^e	6.8 ^e	0.2 ^d	18 ^d	81.8 ^e	8.3 ^d
Means	with	the sar	ne le	tters ar	e not	significantly	differen	t.

Means with the same letters are not significantly different.

* Rh+T₁+T₂= the mixture of Rh.phaseoli+T.viride+T.harzianum, Rh+B₁+B₂ = Rh.phaseoli+ the two

B. subtilis isolates and Rh+ $B_2+T_2=Rh.phaseoli+B.$ subtilis isolate2 + *T. harzianum* isolate2.

REFERENCES

[1.] Abd-El-Moneim, M. L., Tolba, A. F. and Zayan, Sahar A. (2006). Controlling bean soil borne pathogens by compost extract and antagonistic organisms. Minufiya J. Agric. Res., 31(4):759-773.

[2.] Abd-El-Moity, T. H. (1981). Further studies on the biological control of white rot disease of onion. Ph. D. Thesis, Fac. Agric., Minufiya Univ., pp. 135.

[3.] Abd-El-Moity, T. H. (1985). Effect of single and mixture of *Trichoderma harzianum* isolates on controlling three different soil borne pathogens. Egypt. J. Micrbiol., Special Issue.,111-120.

[4.] Abd-El-Moity, T.H; El-Deeb, A.A. and Radwan, I.A. (1991). Biological control of seedling diseases and pod rot of peanuts, under greenhouse and field conditions. Egypt. J. Appl. Sci., 6(1):103-112.

[5.] Abd-El-Khair, H.; Khalifa, R. Kh. M. and Haggag, Karima H.E. (2011).Effect of *Trichoderma* species on damping off diseases incidence, some plant enzymes activity and nutritional status of bean plants. J. of American Science, 7(1):156-167.

[6.] Abdollahi M.; Ommati, F. and Zaker, M. (2012). the in vitro efficacy of *Trichoderma* isolates against *Ptyhium aphanidermatum*, the causal agent of sugar beet root rot. j. Research in Agricultural science, 8(1):79 – 87.

[7.] Abeysinghe, S. (2007). Biological control of *Fusarium solani* f.sp.*phaseoli*, the causal agent of root rot of bean using *Bacillus subtilis* CA32 and *Trichoderma harzianum* RU01. Ruhuna J. of Science. 2: 82-88.

[8.] Ahmadzadeh, M. and Tehrani, A.S. (2009). Evaluation of fluorescent *pseudomonads* for plant growth promotion, antifungal activity against *Rhizoctonia solani* on common bean, and biocontrol potential. Biological Control, 48:101–107.

[9.] Abo-Shosha, Yosra Z. (2006). Biocontrol of *Rhizoctonia solani* damping-off of Bean seedling by *Bacillus* spp. M.Sc. thesis, Plant Pathol. Dept., Fac. Agric., Cairo Univ.

[10.] Akladious S. A. and Abbas, S. M. (2012). Application of *Trichoderma harziunum* T22 as a biofertilizer supporting maize growth. African J. of Biotechnology, 11(35):8672-8683.

[11.] Allen, O.N. (1959). Experiments in soil Bacteriology. 3rd eddition. Burgess Publ. Co. USA. pp.117.

[12.] Attia, M.; Awad, N. M.; Turky, A. S. and Hamed, H. A. (2011). Induction of defense responses in soybean plants against *Macrophomina phaseolina* by some strains of plant growth promoting rhizobacteria. J. of Applied Sci. Res., 1507-1517.

[13.] Balode, A. (2010). Effect of tricodermin, biological product against Botrytis in horticultural crops. Acta Horticulture. 877, 1583-1588.

[14.] Bradford, M. M. (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. Analytical Biochem. 72:248-254.

[15.] Brain, P.W. and Hemming, H. G. (1945). Gliotoxin a fungi static metabolic product of *Trichoderma viride*. Ann. Appl. Biol., 32: 214-220.

[16.] Dowson, W.J. (1957). Plant disease due to bacteria. Second Edition, Cambridge the University Press, London, pp. 23.

[17.] Elad, Y.; Chet,I. and Katan, J. (1980). *Trichoderma harzianum*: A biocontrol agent effective against *Sclerotium rolfsii* and *Rhizoctonia solani*. Phytopathology, 70(2): 119-121.

[18.] Elkoca E.; Turan, M. and Donmez, M. F. (2010). Effect of single, dual and triple inoculations with Bacillus subtilis, Bacillus *megaterium* and *Rhizobium leguminosarum* bv. *Phaseoli* on nodulation, nutrient uptake, yield and yield parameters of common bean (*Phaselus vulgaris* L. cv. Elkoca-05'). J. Plant Nutrition, 33:2104-2119.

[19.] Elsayed, Ayat M. (2013). Control of strawberry fungal diseases under organic agriculture system. Ph.D Thesis, Plant Pathol. Dept., Fac. Agric., Cairo Univ.

[20.] Elsayed, Ayat M.; Shehata,S. T.; Abd-El-Moity, T. H. and Aly, M. M. (2005). Evaluation of single or combined isolates of *Trichoderma harzianum* in different formulations for controlling root rot diseases of strawberry. Annals of Agric. Science, 50(2): 601-611

[21.] Estevez-de-Jensen, C.; Percich, J.A. and Graham, P.H. (2002). Integrated management strategies of bean root rot with *Bacillus subtilis* and *Rhizobium* in Minnesota. Field Crops Research, 74:107-115.

[22.] Ferreira, J.H.S.; Matthee, F.N. and Thomas, A.C. (1991). Biological control of *Eutypa lata* on grapevine by an antagonistic strain of *Bacillus subtilis*. Pytopathology, 81(3): 283-287.

[23.] Graham, P.H., Ranalli, P. (1997). Common bean (*Phaseolus vulgaris* L.). Field Crops Research 53:131-146.

International Journal of Scientific & Engineering Research, Volume 6, Issue 7, July-2015 ISSN 2229-5518

Lorito, M. (2004). Trichoderma species-opportunistic, avirulent Murray 1978, 5.19-228pp.In: Bergey's manual of systemic plant symbionts. Nature Review Microbiology, 2:43-56.

[25.] Karlidag, H.; Esitken, A.; Yildirim, E.; Donmez, M. F. and Turan, M. (2012). Effects of plant growth promoting bacteria on Whitman, W.B.), Springer. 1450 p. yield, growth, leaf water content, membrane permeability, and [32.] Shoresh, M. and Harman, G. E. (2008). The molecular basis ionic composition of strawberry under saline conditions. J. of of shoot responses of maize seedlings to Trichoderma harzianum Plant Nutrition, 34(1):34-45.

and Ghoneim, Soheir, S.H. (1995). A new technique to induce resistance in faba bean against chocolate spot disease. Minufiya J. Phytopathology, 61:1261-1265. Agric. Res., 20(5):1741-1754.

[27.] Mukankusi, C.; Melis, R.; Derera, J.; Laing, M.D. and Buruchara, R.A. (2010). Identification of sources of resistance estimation of reducing sugar and sucrose. J. Am. chem. Soc., 46: to Fusarium root rot among selected common bean lines in 1662-1669. Uganda. J. of Animal & Plant Sciences, 7(3): 876-891.

[28.] Nornai, R. (1982). Formulae for determination of chlorophyllous pigments extracted with N.N. dimethyl formamide, Plant Physiol., 69: 1371-1381.

[29.] Parikka, P.; Kivijarvi, P.; Prokkola, S. and Kemppainen, R. [37.] Unknown, (2013). Foreign trade statistics for the most (2009). Microbiological quality of organic strawberry fruit. Acta Horticulturae, 842: 377-380.

[30.] Peña, P.A.; Steadman, J.R.; Eskridge, K.M.; Urrea, C.A. (2013). Identification of sources of resistance to damping-off and early root/hypocotyl damage from Rhizoctonia solani in common bean (Phaseolus vulgaris L.). Crop Protection, 54: 92-99.

[24.] Harman, G. E.; Howell, C. R.; Viterbo, A.; Chet, I. and [31.] Schleifer, K.H. (2001). Phylum XIII. Firmicutes Gibbons and Bacteriology, second edition. Vol.: 3, The Firmicutes.(Eds. Vos,P.D.; Garrity, G.M.; Jones, D.; Ludwig,W.; Schleifer,K.H. and

T22 inoculation of the root. Plant Physiol., 147(4):2147-2163.

[26.] Mahmoud, Fatma, A.F.; Heweidy, M.A.; Essmat, Nadia E.A., [33.] Simons, T. J. and Ross, A. F. (1971). Changes in metabolism associated with enclosed systemic resistance to tobacco.

> [34.] Thomas, W. and Dutcher, R.A. (1924). The determination of carbohydrate in plants by picric acid reduction method. The

> [35.] Rifai, W. A. (1969). A revision of the genus Trichoderma. Mycological Paper No. 116. Fac. of Pure Since, Univ. of Sheffield, England, 56p.

[36.] Unknown, (2012). Agricultural statistics: publication, 2.

important agricultural commodities, 14.

[38.] Yobo, K. S.; Laing, M. D. and Hunter, C. H. (2011). Effects of single and combined inoculations of selected Trichoderma and Bacillus isolates on growth of dry bean and biological control of Rhizoctonia solani damping-off. Afr. J. Biotechnol., 10(44): 8746-8756.

